



Short Communication

Morphology and Molecular Identification of the *Eimeria* spp. in Domestic Rabbits

Heping Li,¹ Meiyong Shen,¹ Zhijun Hou^{1,3*} and Xiefeng Yin^{2*}

¹College of Wildlife Resources, The Northeast Forestry University, Harbin, China

²College of Environment and Resources, The Inner Mongolia University, Hohhot, China

³Key Laboratory of Wildlife Conservation, China State Forestry Administration, Harbin, P.R. China

ABSTRACT

Although *Eimeria* diseases common in rabbits in China, no information was available on it in Heilongjiang Province. In this study, at least eight *Eimeria* species were detected in domestic rabbits in Harbin in Heilongjiang Province. The diagnosis of *Eimeria* species has been based on the morphology unsporulated oocysts as well as analysis of PCR product obtained by specific primers.

Article Information

Received 28 April 2014

Revised 15 April 2015

Accepted 10 May 2015

Available online 1 January 2016

Authors' Contributions

MS performed PCR. HL and ZH identified *Eimeria* spp. and wrote the article. Data were analyzed by HL and XY.

Key words

Rabbit, *Eimeria*.

Rabbits are mainly infected by monoxenous genus *Eimeria* although they could be infected by the coccidia including the genera *Cryptosporidium*, *Besnoitia*, *Sarcocystis*, *Toxoplasma*, and *Eimeria* (Pakandl, 2009). There are 11 species of rabbit *Eimeria*, which is in agreement with most previous findings, named as *Eimeria stiedai*, *E. intestinalis*, *E. flavescens*, *E. irresidua*, *E. magna*, *E. media*, *E. piriformis*, *E. coecicola*, *E. exigua*, *E. perforans*, and *E. vej dovskyi*. *E. stiedai* develops in the epithelium of bile ducts and others in the intestinal tract (Pakandl, 2009; Oliveira *et al.*, 2011). Among 11 valid *Eimeria* species, *E. stiedai*, *E. intestinalis*, and *E. flavescens* are highly pathogenic in rabbits, especially before the age of 3 months (Pakandl, 2009; Yan *et al.*, 2013; Kvicerova *et al.*, 2008). Most of these *Eimeria* species affect the rabbit production, reduces growth rate and feed conversion, and increases mortality according to their level of pathogenicity (Pakandl, 2009).

Traditional identification of rabbit 11 *Eimeria* spp. was based on their morphological features, such as oocyst size, oocyst residuum (presence or absence), micropyle (conspicuous or inconspicuous), prepatent and patent period, and sporulation time (Kvicerova *et al.*, 2008; Oliveira *et al.*, 2011; Pakandl, 2009). Recently, various molecular differentiation assays, using distinct targets such as ITS1, ITS1-5.8S rRNA-ITS2 and 18S rRNA, have been proposed to identify the rabbit *Eimeria* species (Kvicerova *et al.*, 2008; Oliveira *et al.*, 2011; Yan *et al.*, 2013; Johnson *et al.*, 1991).

In the present study, fecal samples of domestic rabbits suspected of being infected with *Eimeria* species were analyzed based on unsporulated oocyst morphology traits and the molecular method simultaneously.

Materials and methods

Fecal samples were collected from a rabbit farm in Harbin, Northeast of China. All samples were stored at 4°C until analysis. The oocysts were isolated using sedimentation and floatation techniques (Jing *et al.*, 2012). The species were first identified based on the sizes and some other biological features including shape, form index and micropyle of the oocyst (Kvicerova *et al.*, 2008).

DNA extraction was performed according to the methods described by Hou *et al.* (2011). Eleven primer-pairs for PCR were designed to amplify the target fragments following the protocol described by Oliveira (Oliveira *et al.*, 2011). Procedures for avoiding contamination were strictly followed, and negative (no-DNA) controls were included in every experiment. Amplifications were performed in 25µl reaction mixtures consisting of the following: 100 pmol of each primer, 200 µM each deoxynucleoside triphosphate (dNTP), 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl₂ and 2.5 U of Taq DNA polymerase (Takara, Japan) in the appropriate buffer. Reactions were run in a Takara thermocycler using a step-cycle program. Cycling conditions comprised an initial denaturation of DNA at 94°C for 5 min, followed by 35 cycles of 94° for 30 s, each annealing temperature for 30 s, and 72° for 1 min, and a final 10 min 72°C extension step. The PCR products were recovered using the TaKaRa Agarose Gel DNA Purification Kit Ver.2.0 (TaKaRa) and sequenced directly by Invitrogen Company.

* Corresponding authors: houzhihundz@163.com, yin.yxf@gmail.com

Table I.- Morphological characteristics of the rabbit-associated *Eimeria* species.

| Species | Oocysts shape | Oocysts size (µm) | Index | Micropyle |
|---|--------------------------|-------------------|-------|--------------------------------------|
| <i>E. vej dovskiyi</i> (or <i>E. coecicola</i>) | elongate-ovoid | 29.11×15.55 | 1.87 | Conspicuous |
| <i>E. intestinalis</i> (or <i>E. piriformis</i>) | piriform | 26.72×16.96 | 1.58 | Conspicuous; on the narrow polar |
| <i>E. flavescens</i> | piriform | 24.82×16.05 | 1.55 | Conspicuous; on the widest polar |
| <i>E. magna</i> | ovoid | 33.24×17.92 | 1.85 | Conspicuous; outside the oocyst wall |
| <i>E. media</i> | ellipsoid-ovoid | 30.62×18.35 | 1.67 | Conspicuous |
| <i>E. stiedai</i> | ellipsoid | 30.14×18.32 | 1.65 | Conspicuous |
| <i>E. perforans</i> | ellipsoid-subrectangular | 20.26×12.09 | 1.68 | inconspicuous |
| <i>E. exigua</i> | spherical-sub spherical | 13.66×12.82 | 1.07 | inconspicuous |

Table II.- The nucleotide sequence alignment results of *Eimeria* spp. in BLAST search and the Genbank Accession numbers.

| Accession No. | Identities | Expect | The nucleotides and sequence numbers | | |
|--|---------------|--------|--------------------------------------|--------------------|--------------------|
| <i>E. vej dovskiyi</i> HRB (1-164) HM768891.1 (172-337) | 99%(163/164) | 1e-47 | A(107) G(278) | deletion C(281) | deletion T(282) |
| <i>E. piriformis</i> -HRB(1-289) HM768889.1 (102-390) | 100%(289/289) | 2e-148 | \ | \ | \ |
| <i>E. flavescens</i> -HRB(1-199) HM768883.1 (196-394) | 99%(198/199) | 7e-97 | A(110) T(305) | \ | \ |
| <i>E. magna</i> -HRB (1-213) JQ071391.1 (173-385) | 99%(211/213) | 6e-103 | A(15) G(187) | T(193) C(365) | |
| <i>E. media</i> -HRB (1-152) JQ071392.1 (259-410) | 100%(152/152) | 2e-72 | \ | \ | \ |
| <i>E. stiedai</i> -HRB (1-217) JQ328190.1 (127-343) | 99% (215/217) | 4e-105 | A(101) T(227) | A(165) G(291) | \ |

Results and discussion

There were at least eight species detected in this study, including *Eimeria vej dovskiyi* (or *E. coecicola*), *E. intestinalis* (or *E. piriformis*), *E. flavescens*, *E. magna*, *E. media*, *E. stiedai*, *E. perforans*, and *E. exigua* (Fig. 1, Table I).

Six *Eimeria* spp. species were detected by PCR method, and they were *E. vej dovskiyi* (164bp), *E. piriformis* (289bp), *E. flavescens* (199bp), *E. magna* (213bp), *E. media* (152bp), and *E. stiedai* (217bp). The sequences obtained in this study by amplifying the *Eimeria* oocysts ITS1 was submitted to a BLAST search for similarity (www.ncbi.nlm.nih.gov/blast), and the identities were 99% to 100% (Table II).

In the present study, at least eight *Eimeria* spp. were based on unsporulated oocysts morphology characters, but only six species were detected by PCR assays. The unsporulated morphology characteristics between *E. vej dovskiyi* and *E. coecicola*, *E. piriformis* and

E. intestinalis were similar, so it was impossible to differentiate them only on the basis of unsporulated morphology features. Other *Eimeria* spp. had either different oocyst size and oocyst shape, or micropyle, and could be differentiated from each other by those unsporulated biology traits excepting the *E. perforans* and *E. exigua*.

The classic diagnostic method of the rabbit *Eimeria* spp. is based on morphology which needs to sporulate examining for days before inspecting. The molecular method accurate and sensitive was time-consuming. This problem could be resolved by detecting the *Eimeria* spp. infecting rabbits based on their unsporulated morphology characteristics such as oocyst shape, oocyst size, and micropyle. Although it was not accurate to distinguish *E. vej dovskiyi* from *E. coecicola*, *E. piriformis* from *E. intestinalis*, or even others species, this method could be used to identify most of the *Eimeria* species in infected rabbits. The highly pathogenic species

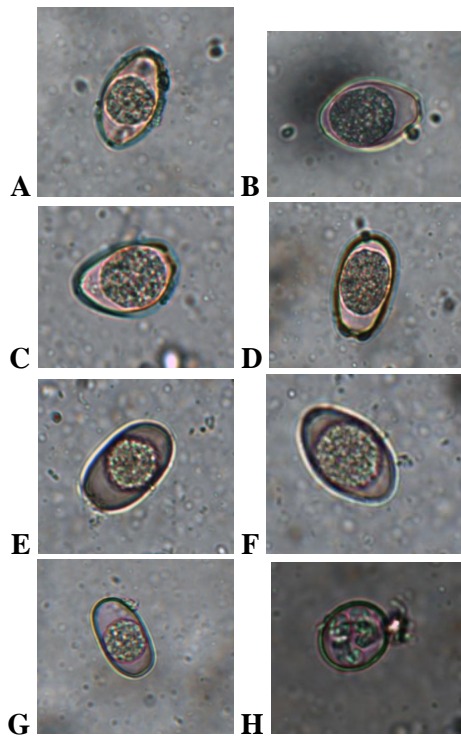


Fig. 1. The *Eimeria* species detected in the rabbits in this study ($\times 400$). A, *Eimeria vej dovskyi* (or *E. coecicola*); B, *E. intestinalis* (or *E. piriformis*); C, *E. flavescens*; D, *E. magna*; E, *E. media*; F, *E. stiedai*; G, *E. perforans*; H, *E. exigua*.

(*E. stiedai*, *E. flavescens*, and *E. intestinalis*) and wild pathogenic species (*E. magna*, *E. media*, and *E. piriformis*) can be distinguished based on the unsporulated morphological features if permitted to take the *Eimeria* spp oocyst with piriform shape as intestinalis. Enough information for controlling the disease was inferred timely based on these features because there were not many drugs to be selected for treatment (Jing *et al.*, 2012), even though all *Eimeria* spp. were differentiated accurately. Moreover, most drugs have similar action on most *Eimeria* species. Therefore, diagnosing the *Eimeria* infection based on the

unsporulated oocyst morphology was practical for small rabbit farms for controlling and preventing the rabbit *Eimeria* diseases.

In this study, the *E. perforans* and *E. exigua* were identified based on the morphological characteristics, but could not be detected by the PCR methods

To our knowledge, this is the first report on prevalence of *E. vej dovskyi* in China. Jing *et al.* (2012) reported that 8 species prevailed in the Northeast China except *E. vej dovskyi*, *E. stiedai*, and *E. exigua*. The reason for this might be because they did not collect the samples in Heilongjiang province, one main area of Northeast China.

In conclusion, we found that at least eight *Eimeria* species prevailed in Harbin, and had developed a practical method to diagnose the rabbit *Eimeria* disease timely for small rabbit farms.

Acknowledgements

This study was supported by the Fundamental Research Funds for the Central Universities (No.2572015CA20, DL13EA01) and The National Nature Science Found (21107040).

Conflict of interest statement

Authors have no conflict of interest to declare.

References

- Johnson, A. M., Ellis, R.F., O'Donoghue, P. J. and Baverstock, P. R., 1991. *Syst. Parasitol.*, **18**:1-8.
- Jing, F., Yin, G., Liu, X., Suo, X. and Qin, Y., 2012. *Parasitol. Res.*, **110**:1495-1500.
- Hou, Z., Xing, M., Chai, H. and Hua, Y., 2011. *Pakistan J. Zool.*, **43**: 505-510.
- Kvicerova, J., Pakandl, M. and Hypsa, V., 2008. *Parasitology*, **135**: 443-452.
- Oliveira, U.C., Fraga, J.S., Licois, D., Pakandl, M. and Gruber, A., 2011. *Vet. Parasitol.*, **176**: 275-280.
- Pakandl, M., 2009. *Folia Parasitol.*, **56**: 153-156.
- Yan, W., Wang, W., Wang, T., Suo, X., Qian, W., Wang, S. and Fan, D., 2013. *Vet. Parasitol.*, **193**: 284-288.